

# miRStar™ Human Cancer Focus miRNA PCR Array

Cat#: AS-MR-001

## Instruction Manual version 1.0

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## Product Summary

### Kit components

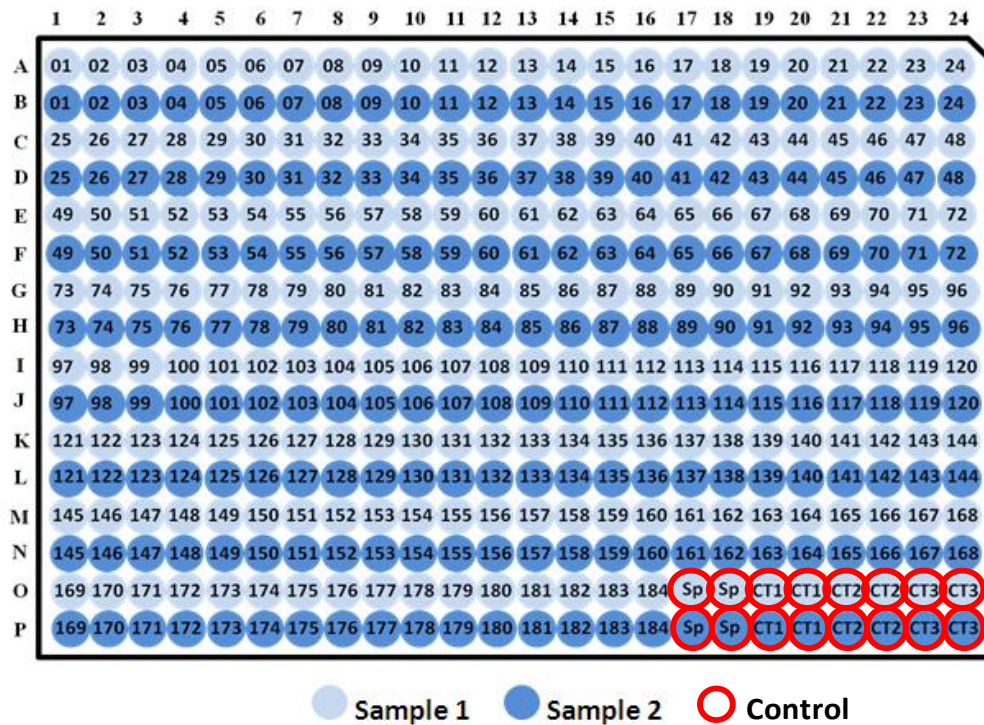
Catalog Number	Contents	Storage
AS-MR-001	miRStar™ Human Cancer Focus miRNA PCR Array, dried down assays in 384-well plate	-20°C

### Description

Arraystar's miRStar™ Human Cancer Focus microRNA PCR Array contains two identical sets of 184 miRNAs most relevant to cancer. Two individual samples, for example tumor vs. peritumoral or biological replicates, can be profiled in parallel. To ensure high data quality, the panel includes 8 miRNA reference sets to better quantify and normalize the qPCR data. cDNA synthesis and PCR efficiency are evaluated by using the synthetic cel-miR-39-3p as the Spike-in RNA control. The array is a powerful tool to conveniently and quickly analyze the expression levels of miRNAs most relevant to cancers, which is valuable for cancer biology research and cancer biomarker discovery.

### Array Layout

The cancer-associated miRNAs for each sample are in the alternate rows (shaded in light or darker colors for Sample 1 and Sample 2). The control assays are circled in red.



**Figure 1.** The array layout for miRStar Human Cancer Focus miRNA PCR Array.

- #01 through #184                      184 cancer-associated miRNAs.
  
- O17 and O18                              Spike-in Control (**SP**) in duplicate, to evaluate cDNA synthesis and PCR efficiency.
- P17 and P18
  
- O19 through O24                        Three small nuclear or small nucleolar RNAs in duplicates, RNU6-2-F (**CT1**), SNORD43-F (**CT2**), and SNORD95-F (**CT3**), to normalize qPCR data for the miRNAs.
- P19 through P24
  
- #51, #66, #144, #146, #177      Five housekeeping miRNAs as the internal quantification controls or reference genes: hsa-miR-16-5p(#51, Well E03 and F03), hsa-miR-191-5p(#66, Well E18 and F18), has-miR-423-3p(#144, Well K24 and L24),

hsa-miR-425-5p(#146, Well M02 and N02), and hsa-miR-93-5p(#177, Well O09 and P09).

### Description of Control Assays

There are three types of control assays built in the miRStar Human Cancer Focus miRNA PCR Array 384HC. Each control assay is in duplicate. Their uses and meanings are explained below.

- **SP** (Spike-in control): An RNA spike-in control is added in the RNA sample during the first-strand cDNA synthesis (Protocol Step A2). The SP control assay indicates the overall success and the efficiency of the reactions beginning from the adaptor ligation, cDNA synthesis to the final qPCR. Any problem(s) in these steps will result in a failed or compromised SP outcome.
- **CT** (miRNA Control Reference): Three stably expressed small nuclear or small nucleolar RNA genes RNU6-2-F (**CT1**), SNORD43-F (**CT2**), and SNORD95-F (**CT3**) are included in the array as the quantification references for miRNA. Additionally,
- **Housekeeping miRNA genes**: Five housekeeping miRNAs, namely, hsa-miR-16-5p(#51, Well E03 and F03), hsa-miR-191-5p(#66, Well E18 and F18), has-miR-423-3p(#144, Well K24 and L24), hsa-miR-425-5p(#146, Well M02 and N02), and hsa-miR-93-5p(#177, Well O09 and P09), can also serve as the internal quantification or reference controls.

### List of miRNAs and controls

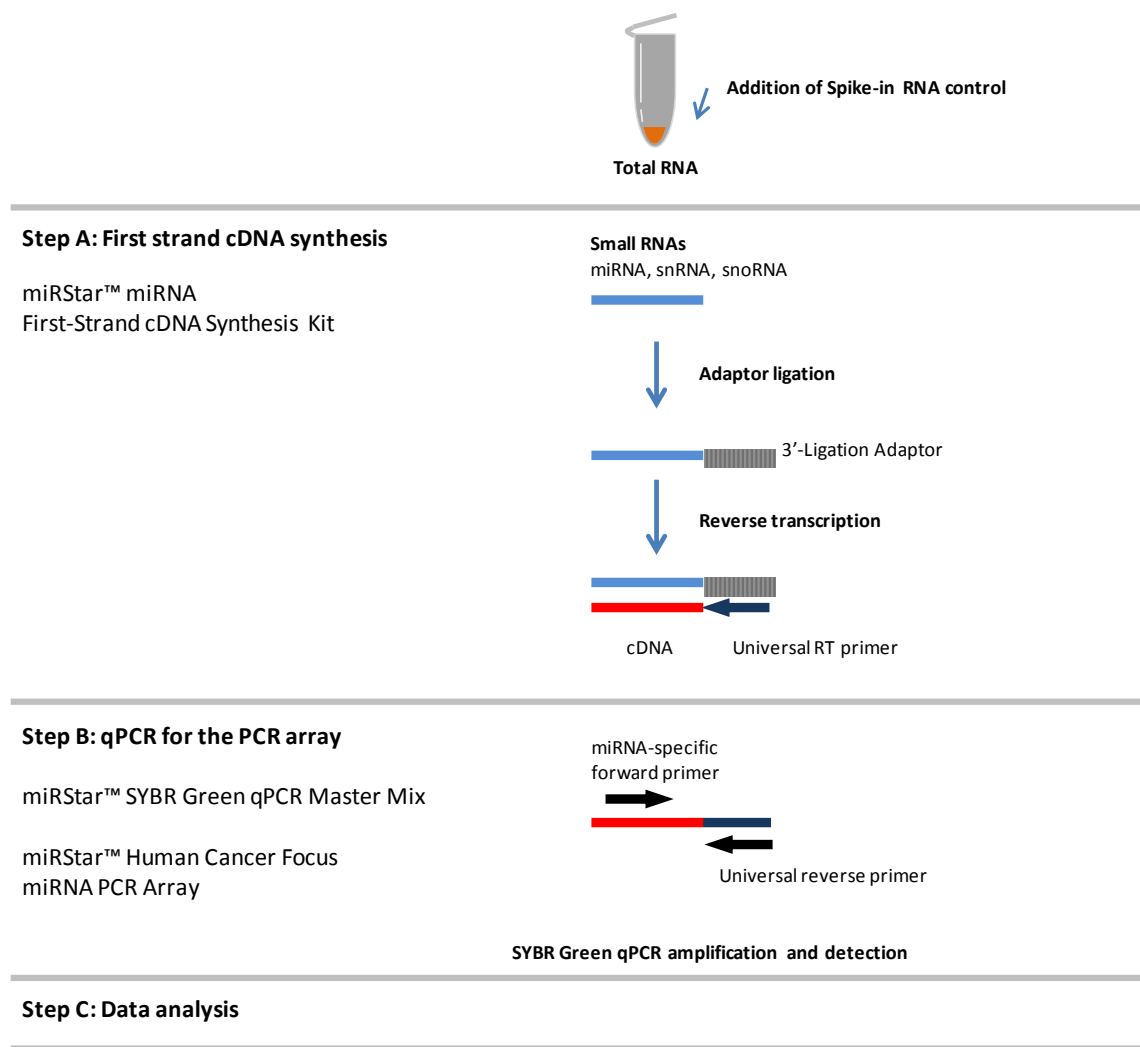
Control RNAs are outlined in red.

A01	B01	E01	F01	I01	J01	M01	N01	miR-424-5p
A02	B02	E02	F02	I02	J02	M02	N02	miR-425-5p
A03	B03	E03	F03	I03	J03	M03	N03	miR-429
A04	B04	E04	F04	I04	J04	M04	N04	miR-451a
A05	B05	E05	F05	I05	J05	M05	N05	miR-486-5p
A06	B06	E06	F06	I06	J06	M06	N06	miR-495-3p
A07	B07	E07	F07	I07	J07	M07	N07	miR-497-5p
A08	B08	E08	F08	I08	J08	M08	N08	miR-501-5p
A09	B09	E09	F09	I09	J09	M09	N09	miR-502-3p
A10	B10	E10	F10	I10	J10	M10	N10	miR-505-3p
A11	B11	E11	F11	I11	J11	M11	N11	miR-517a-3p
A12	B12	E12	F12	I12	J12	M12	N12	miR-518a-3p
A13	B13	E13	F13	I13	J13	M13	N13	miR-518b
A14	B14	E14	F14	I14	J14	M14	N14	miR-518c-3p
A15	B15	E15	F15	I15	J15	M15	N15	miR-518e-3p
A16	B16	E16	F16	I16	J16	M16	N16	miR-518f-3p
A17	B17	E17	F17	I17	J17	M17	N17	miR-519d
A18	B18	E18	F18	I18	J18	M18	N18	miR-524-5p
A19	B19	E19	F19	I19	J19	M19	N19	miR-532-5p
A20	B20	E20	F20	I20	J20	M20	N20	miR-539-5p
A21	B21	E21	F21	I21	J21	M21	N21	miR-584-5p
A22	B22	E22	F22	I22	J22	M22	N22	miR-617
A23	B23	E23	F23	I23	J23	M23	N23	miR-629-5p
A24	B24	E24	F24	I24	J24	M24	N24	miR-652-3p
C01	D01	G01	H01	I01	L01	O01	P01	miR-7-5p
C02	D02	G02	H02	I02	L02	O02	P02	miR-744-5p
C03	D03	G03	H03	I03	L03	O03	P03	miR-777-5p
C04	D04	G04	H04	I04	L04	O04	P04	miR-885-5p
C05	D05	G05	H05	I05	L05	O05	P05	miR-886-3p
C06	D06	G06	H06	I06	L06	O06	P06	miR-9-5p
C07	D07	G07	H07	I07	L07	O07	P07	miR-92a-3p
C08	D08	G08	H08	I08	L08	O08	P08	miR-92b-3p
C09	D09	G09	H09	I09	L09	O09	P09	miR-93-5p
C10	D10	G10	H10	I10	L10	O10	P10	miR-93-3p
C11	D11	G11	H11	I11	L11	O11	P11	miR-96-5p
C12	D12	G12	H12	I12	L12	O12	P12	miR-96-3p
C13	D13	G13	H13	I13	L13	O13	P13	miR-98-5p
C14	D14	G14	H14	I14	L14	O14	P14	miR-99a-3p
C15	D15	G15	H15	I15	L15	O15	P15	miR-99a-5p
C16	D16	G16	H16	I16	L16	O16	P16	miR-99b-5p
C17	D17	G17	H17	I17	L17	O17	P17	C. e miR-39
C18	D18	G18	H18	I18	L18	O18	P18	C. e miR-39
C19	D19	G19	H19	I19	L19	O19	P19	RNU6-2-F
C20	D20	G20	H20	I20	L20	O20	P20	RNU6-2-F
C21	D21	G21	H21	I21	L21	O21	P21	SNORD43-F
C22	D22	G22	H22	I22	L22	O22	P22	SNORD43-F
C23	D23	G23	H23	I23	L23	O23	P23	SNORD95-F
C24	D24	G24	H24	I24	L24	O24	P24	SNORD95-F

# Protocol

## Workflow Overview

A miRStar Human Cancer Focus miRNA PCR Array experiment consists of several major steps in a workflow shown in Figure 2.



**Figure 2.** Workflow overview of miRStar™ Human Cancer Focus miRNA PCR Array experiment.

### Step A. First-strand cDNA synthesis

Total RNA samples should be extracted by a method that can recover small RNA fraction, for example, TRIzol® Reagent method.

High quality cDNA synthesis is vital for the following qPCR performance. We highly recommend Arraystar miRNA First-Strand cDNA Synthesis Kit (Cat# AS-MR-004), which is fully compatible with and is specifically optimized for miRStar Human Cancer Focus miRNA PCR Array. Please refer to the Instruction Manual of the Kit for its use.

1. Dilute the 3' Ligation Adapter from the Kit with RNase-free water. The dilution factor is 1/10 for 10 - 500 ng or 1/3 for 0.5 - 2 µg of the starting total RNA. Use the same amount of total RNA for each sample in the experiment.
2. Set up the adaptor ligation reaction in a 200 µL PCR tube using the following components for each sample:

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4.5 µL	Total RNA in nuclease-free water
1.0 µL	diluted 3' Ligation Adapter
1.0 µL	RNA Spike-in

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6.5 µL	total volume
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3. Incubate in a thermal cycler at 70°C for 2min; chill on ice immediately.
4. Add the following reagents and mix well. The final volume will be 10 µL.

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2.0 µL	5×Ligase Reaction Mix
1.0 µL	RNA ligase
0.5 µL	RNase Inhibitor

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10.0 µL	final volume
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5. Incubate at 22°C for 60 min; 72°C for 2 min; and on ice for 2 min.

6. For reverse transcription, add 1  $\mu\text{L}$  Universal RT Primer Mix, mix gently.
7. Incubate at 65°C for 2 min; place on ice for at least 2 min.
8. Prepare Reverse Transcription Master Mix and add 10  $\mu\text{L}$  to each sample above.

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8.5 $\mu\text{L}$	RT Reaction Master Mix
0.5 $\mu\text{L}$	RNase Inhibitor
1.0 $\mu\text{L}$	MMLV Reverse Transcriptase

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10.0 $\mu\text{L}$	total volume per sample
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9. Incubate at 42°C for 60 min; inactivate the reaction at 85°C for 5 min.

### Step B. Perform qPCR for the PCR array

1. Dilute the cDNA in nuclease free water. If Arraystar miRNA First-Strand cDNA Synthesis Kit (Cat# AS-MR-004) is used for the cDNA synthesis with 10 ng - 2.0  $\mu\text{g}$  total RNA sample as the starting material, dilute the cDNA product 1/80 in water. The diluted material is used as the qPCR template.
2. Use Arraystar SYBR Green Real-Time Quantitative PCR Master Mix to prepare qPCR Master Mix for each sample per qPCR well. There are total 384 reactions in a 384-well qPCR array plate, 192 wells for miRNA and 192 wells for mRNA (Figure 1). Add some extra reactions as needed by the liquid handling operation. Multiply this number with the individual amounts of the components in the table below and prepare a qPCR Mix.

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5.0 $\mu\text{L}$	SYBR Green Master Mix
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1.5 µL	Diluted cDNA template
3.5 µL	ddH <sub>2</sub> O
<hr/>	
10.0 µL	total volume per well

3. Dispense 10 µl of the Mix uniformly to each well on the qPCR array plate.
4. Run the qPCR using the following program. Consult the instructions for the instrument for details.

Cycles	Temperature	Time
1	95 °C	10 minutes
40	95 °C	10 seconds
	55-65 °C	1 minute
Melting curve analysis		

### Step C. Data analysis

1. Calculate the  $\Delta Ct$  for each miRNA:

$$\Delta Ct_{\text{miRNA}} = Ct_{\text{miRNA}} - \text{average}(Ct_{\text{control}})$$

Where  $Ct_{\text{control}}$  are the values taken from one or more duplicates of the miRNA references (**CT**).

If no particular reference gene(s) are designated as the quantification reference, all the CTs can be averaged and used in the above formula, but only if the difference between the averaged values is less than 1 cycle when comparing the two groups.

2. Calculate the  $\Delta\Delta Ct$  between two samples or groups for a gene:

$$\Delta\Delta Ct = \Delta Ct_{\text{sample2}} - \Delta Ct_{\text{sample1}}, \text{ or}$$

$$\Delta\Delta Ct = \Delta Ct_{\text{group2}} - \Delta Ct_{\text{group1}}$$

Where sample1 or group1 is the control and sample2 or group2 is the experimental.

3. Calculate the fold change from group 1 to group 2 for a gene as:

$$\text{fold change} = 2^{-\Delta\Delta Ct}$$

OPTIONAL: If the fold-change is greater than 1, the result may be reported as a fold up-regulation. If the fold-change is less than 1, the negative reciprocal may be reported as a fold down-regulation.



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